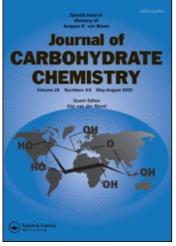
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# Synthesis of the Tetrasaccharide Repeating Unit of the Antigen from *Escherichia coli* O126 as Its Methyl Glycoside

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# SYNTHESIS OF THE TETRASACCHARIDE REPEATING UNIT OF THE ANTIGEN FROM ESCHERICHIA COLI O126 AS ITS

METHYL GLYCOSIDE

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#### ABSTRACT

The tetrasaccharide repeating unit (17) of the antigen from *E. coli* O126 has been synthesized as its methyl glycoside by sequential addition of monosaccharide derivatives. The formation of the  $\beta$ -mannosidic linkage was achieved by Swern oxidation of the glucose derivative followed by reduction of the product with sodium borohydride.

#### **INTRODUCTION**

Escherichia coli is a complex group of gram-negative bacteria and many of its serotypes are important human pathogens causing extraintestinal and intestinal infections.<sup>1-3</sup> The serological classification of *E.coli* is mainly based on the nature of O-antigens, i.e., the O-specific polysaccharide part of the lipopolysaccharide (LPS), which is the major outer membrane component of the bacteria.<sup>4</sup> It was reported that natural immunity to gram-negative bacteria is often provided by antibodies that recognize LPS.<sup>5</sup> Such recognition involves a part of the structure of the polysaccharide and these substructures may ultimately provide a tool to study the structure-function relationship.<sup>6</sup>

The enteropathogenic strain of *E.coli* O126 is known to be associated with infantile diarrhea and the structure of the O-specific polysaccharide from this strain has already been reported.<sup>7</sup> In view of increasing drug resistance of the pathogenic bacteria and potential importance of artificial antigen, chemical synthesis of the repeating unit (I) of the antigen has gained considerable interest. This will help in designing synthetic antigens for precise diagnosis and protection. We have already published the synthesis of a trisaccharide<sup>8</sup> related to *E.coli* O126 antigen. In this communication, we report the chemical synthesis of its tetrasaccharide repeating unit.

$$\rightarrow 3) \cdot \beta \cdot D \cdot Glcp NAc \cdot (1 \rightarrow 2) \cdot \beta \cdot D \cdot Manp \cdot (1 \rightarrow 3) \cdot \alpha \cdot D \cdot Galp \cdot (1 \rightarrow 2)$$

$$\uparrow$$

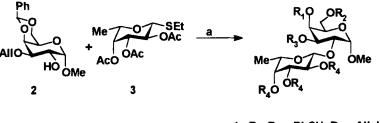
$$I$$

$$I$$

$$\beta \cdot L \cdot Fucp$$

#### **RESULTS AND DISCUSSION**

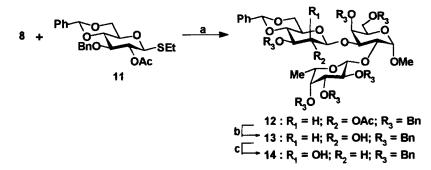
Methyl 3-O-allyl- $\alpha$ -D-galactopyranoside (1),<sup>9</sup> prepared from D-galactose was allowed to react with benzaldehyde dimethyl acetal in the presence of *p*toluenesulfonic acid to give crystalline 2 in 76% yield. The acceptor 2 was allowed to react with the thioglycoside 3<sup>10</sup> in the presence of NIS/TfOH as promoter<sup>11</sup> to afford the disaccharide 4 in 72% yield. Removal of the benzylidene group of 4 with 85% acetic acid<sup>12</sup> followed by Zemplén deacetylation<sup>13</sup> and conventional benzylation<sup>14</sup> of the product gave the pentabenzyl derivative 7 in 70% overall yield. The disaccharide 7 was deallylated<sup>15</sup> with palladium(II) chloride in methanol to give the disaccharide acceptor 8 in 90% yield (Scheme 1).



 $b \longrightarrow 4: R_1, R_2 = PhCH; R_3 = Allyl; R_4 = Ac$   $b \longrightarrow 5: R_1 = R_2 = H; R_3 = Allyl; R_4 = Ac$   $c \longrightarrow 6: R_1 = R_2 = R_4 = H; R_3 = Allyl$   $d \longrightarrow 7: R_1 = R_2 = R_4 = Bn; R_3 = Allyl$   $e \longrightarrow 8: R_1 = R_2 = R_4 = Bn; R_3 = H$ 

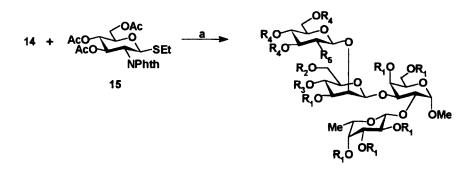
Scheme 1: (a) NIS/TfOH/CH<sub>2</sub>Cl<sub>2</sub>/MS-4Å/-20 °C/25 min; (b) 85% AcOH/ 80 °C/2 h; (c) 0.05 M MeONa/MeOH/r t /3 h; (d) BnBr/NaH/DMF/r t/6 h; (e) PdCl<sub>2</sub>/MeOH/r t/4 h.

In a separate experiment, ethyl 4,6-O-benzylidene-1-thio- $\beta$ -D-glucopyranoside (9)<sup>16</sup> was selectively 3-O-benzylated<sup>17</sup> using dibutyltin oxide and cesium fluoride giving 10, which was then acetylated<sup>18</sup> to afford 11. The disaccharide acceptor 8 was allowed to react with 11 in the presence of NIS/TfOH as promoter<sup>11</sup> to give the trisaccharide 12 in 69% yield. Deacetylation<sup>13</sup> of 12 gave crystalline 13 with a hydroxyl group at the 2" position. Swern oxidation<sup>19</sup> of 13 using dimethyl sulfoxide-acetic anhydride followed by reduction of the product with sodium borohydride afforded the acceptor 14 in 62% yield (Scheme 2).



Scheme 2 : (a) NIS/TfOH/CH<sub>2</sub>Cl<sub>2</sub>/MS-4Å/-25 °C/20 min; (b) 0.05 M MeONa/ MeOH/r t//8 h;(c) (I) 2:1 Me<sub>2</sub>SO-Ac<sub>2</sub>O, (ii) NaBH<sub>4</sub>/1:1 MeOH-CH<sub>2</sub>Cl<sub>2</sub>/r t/6 h.

The trisaccharide acceptor 14 was then allowed to react with ethyl 3,4,6-tri-O-acetyl-2-deoxy-2-phthalimido-1-thio- $\beta$ -D-glucopyranoside (15)<sup>20</sup> in the presence of NIS/TfOH as promoter<sup>11</sup> to give the tetrasaccharide derivative 16 in 64% yield. The <sup>1</sup>H NMR spectrum of 16 gave signals at  $\delta$  4.78, 4.67, 4.62 and 5.53 ppm for H-1, H-1', H-1" and H-1" respectively. Dephthaloylation of 16 with hydrazine hydrate<sup>21</sup> followed by N-acetylation and hydrogenolysis of the product afforded the target tetrasaccharide repeating unit as its methyl glycoside (17) in 67% yield (Scheme 3). The <sup>1</sup>H NMR spectrum of 17 showed anomeric signals for one  $\alpha$ -D-galactopyranosyl ( $\delta$  4.70), one  $\beta$ -L-fucopyranosyl ( $\delta$  4.30), one  $\beta$ -D-mannopyranosyl ( $\delta$  4.16) and one 2-acetamido-2-deoxy- $\beta$ -D-glucopyranosyl ( $\delta$  4.83) moiety, together with a peak for a methyl glycoside anomeric proton.



16 :  $R_1 = Bn; R_2, R_3 = PhCH; R_4 = Ac; R_5 = NPhth$  $b \longrightarrow 17 : R_1 = R_2 = R_3 = R_4 = H; R_5 = NHAc$ 

Scheme 3 : (a) NIS/TfOH/CH<sub>2</sub>Cl<sub>2</sub>/MS-4Å/-20 °C/30 min; (b) (i) 10% Pd-C/H<sub>2</sub>/AcOH/r t/48 h, (ii) NH<sub>2</sub>NH<sub>2</sub>.H<sub>2</sub>O/EtOH/90 °C/2 h, (iii) Pyridine/Ac<sub>2</sub>O/r t/4 h, (iv) 0.05 M MeONa/MeOH/4 h.

#### **EXPERIMENTAL**

General Procedures. All reactions were monitored by TLC on Silica Gel G (E. Merck, India). Column chromatography was performed using silica gel (SRL, India) and all concentrations were conducted below 50 °C unless stated otherwise. Optical rotations were measured at 24 °C with a Perkin-Elmer 241 MC polarimeter. The <sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded with Bruker DPX 300 instrument using  $CDCl_3$  as solvent and TMS as the internal standard unless stated otherwise.

Methyl 3-O-Allyl-4,6-O-benzylidene- $\alpha$ -D-galactopyranoside (2). To a solution of 1 (6.14 g, 26.24 mmol) in CH<sub>3</sub>CN (60 mL), were added benzaldehyde dimethyl acetal (6 mL, 39.5 mmol), *p*-TsOH (150 mg) and MS-3Å (7 g) and the mixture was then stirred at room temperature for 16 h. The reaction was quenched with Et<sub>3</sub>N (0.5 mL), and the reaction mixture was filtered through a Celite bed and concentrated. The crude product was crystallized from EtOH to give 2 (6.4 g, 76%): mp 158-159 °C; [ $\alpha$ ]<sub>D</sub> +191.8° (*c* 1.09, CHCl<sub>3</sub>); <sup>1</sup>H NMR  $\delta$  3,43 (s, 3H, OCH<sub>3</sub>), 4.82 (d, 1H, J=3.2 Hz, H-1), 5.50 (s, 1H, PhCH), 6.0 (m, 1H, CH<sub>2</sub>CH=CH<sub>2</sub>), 7.30-7.56 (m, 5H, aromatic protons).

Anal. Calcd for C<sub>17</sub>H<sub>22</sub>O<sub>6</sub> : C, 63.34; H, 6.88. Found : 63.13; H, 7.08.

Methyl 2,3,4-Tri-O-acetyl- $\beta$ -L-fucopyranosyl- $(1\rightarrow 2)$ -3-O-allyl-4,6-O-benzylidene- $\alpha$ -D-galactopyranoside 3 (1.95 g, 5.84 mmol), 2 (1.26 g, 3.9 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (40 mL), MS-4Å (6 g) was added and the mixture was stirred under Ar at room temperature for 18 h. NIS (1.72 g, 7.65 mmol) was then added, the mixture was allowed to cool to -25 °C and TfOH (67 $\mu$ L, 0.76 mmol) was injected into the cooled reaction mixture. Stirring was continued at -20 °C for 20 min, the reaction mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub> and filtered through a Celite bed. The filtrate was washed with 10% Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub>, aq NaHCO<sub>3</sub>, water, dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated to a syrupy product. Column chromatography using 4:1 toluene-Et<sub>2</sub>O then gave 4 (2.9 g, 72%); [ $\alpha$ ]<sub>D</sub> + 50.3° (c 0.9, CHCl<sub>3</sub>); <sup>1</sup>H NMR  $\delta$  1.18 (d, 3H, J=6.1 Hz, CCH<sub>3</sub>), 1.97, 2.06 and 2.14 (3s, 9H, 3COCH<sub>3</sub>), 3.40 (s, 3H, OCH<sub>3</sub>), 4.65 (d, 1H, J=7.8 Hz, H-1'), 4.81 (d, 1H, J=3.3 Hz, H-1), 5.54 (s, 1H, PhCH), 5.98 (m, 1H, CH<sub>2</sub>-CH=CH<sub>2</sub>), 7.26-7.54 (m, 5H, aromatic protons).

Anal. Calcd for C<sub>29</sub>H<sub>38</sub>O<sub>13</sub> : C, 58.58; H, 6.40. Found: C, 58.40; H, 6.63.

Methyl 2,3,4-Tri-O-benzyl- $\beta$ -L-fucopyranosyl- $(1 \rightarrow 2)$ -3-O-allyl-4,6di-O-benzyl- $\alpha$ -D-galactopyranoside (7). A solution of 4 (2.8 g, 4.71 mmol) in 85% AcOH (50 mL) was stirred at 90 °C for 3 h. Removal of the solvent gave 5 (2.14 g; 90%) which was treated with 0.05 M MeONa in MeOH (40 mL). After 2 h, the solution was made neutral by the addition of Amberlite IR-120 (H<sup>+</sup>) resin, filtered and the solvent was evaporated to provide 6 (2.1 g, 96%). To a solution of 6 (1.6 g, 4.21 mmol) in DMF (30 mL) were added NaH (50% oil coated, 2.0 g, 41.6 mmol) and BnBr (3.75 mL, 31.5 mmol), and the mixture was stirred at room temperature for 6 h. MeOH (3 mL) was then added to destroy the excess reagents, the reaction mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub> (150 mL) and washed with water, dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated to give a crude syrup. Column chromatography using 10:1 toluene-Et<sub>2</sub>O gave 7 (2.44 g, 70%):  $[\alpha]_D$  + 1.05° (c 0.9, CHCl<sub>3</sub>); <sup>1</sup>H NMR  $\delta$  1.21 (d, 3H, J=6.2 Hz, CCH<sub>3</sub>), 3.30 (s, 3H, OCH<sub>3</sub>), 4.66 (d, 1H, J=7.2 Hz, H-1'), 4.88 (d, 1H, J=3.3 Hz, H-1), 5.96 (m, 1H, CH<sub>2</sub>CH=CH<sub>2</sub>), 7.19-7.40 (m, 25H, aromatic protons).

Anal. Calcd for C<sub>51</sub>H<sub>58</sub>O<sub>10</sub> : C,73.73; H, 6.98. Found: C, 73.54; H, 7.19.

Methyl 2,3,4-Tri-O-benzyl-β-L-fucopyranosyl- $(1\rightarrow 2)$ -4,6-di-Obenzyl-α-D-galactopyranoside (8). A mixture of 7 (2.1 g, 2.53 mmol) and PdCl<sub>2</sub> (149 mg, 0.84 mmol) in dry MeOH (30 mL) was stirred at room temperature for 4 h. The reaction mixture was filtered through a celite bed and the filtrate was concentrated. Column chromatography of the crude product using 8:1 toluene-Et<sub>2</sub>O gave pure 8 (1.8 g, 90%): [α]<sub>D</sub> +13.6° (c 0.8, CHCl<sub>3</sub>); <sup>1</sup>H NMR δ 1.23 (d, 3H, J=6.3 Hz, CCH<sub>3</sub>), 3.31 (s, 3H, OCH<sub>3</sub>), 4.66 (d, 1H, J=7.5 Hz, H-1'), 4.83 (d, 1H, J=3.6 Hz, H-1), 7.24-7.37 (m, 25H, aromatic protons).

Anal. Calcd for C<sub>48</sub>H<sub>54</sub>O<sub>10</sub> : C, 72.91; H, 6.83. Found: C, 72.76; H, 7.05.

Ethyl 3-O-Benzyl-4,6-O-benzylidene-1-thio- $\beta$ -D-glucopyranoside (10). A mixture of ethyl 4,6-O-benzylidene-1-thio- $\beta$ -D-glucopyranoside 9 (2.73 g, 8.75 mmol), dibutyltin oxide (2.6 g, 10.44 mmol) in MeOH (50 mL) was refluxed. After 1 h the reaction mixture became clear, and the solution was concentrated. The residue was dried under vacuum and dissolved in DMF (30 mL). To this solution, predried CsF (1.6 g, 10.53 mmol) and BnBr (1.6 mL, 13.47 mmol) were added and the mixture was stirred at room temperature for 20 h. The reaction mixture was concentrated, the residue was partitioned between CH<sub>2</sub>Cl<sub>2</sub> and water, and the organic layer was dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated to a solid mass. Column chromatography using 6:1 toluene-Et<sub>2</sub>O gave pure 10 (3.0 g, 85%), which was crystallized from EtOH: mp 145-146 °C;  $[\alpha]_D$  -54.5° (c 1.0, CHCl<sub>3</sub>); <sup>1</sup>H NMR  $\delta$  1.30 (t, 3H, SCH<sub>2</sub>CH<sub>3</sub>), 2.74 (q, 2H, SCH<sub>2</sub>CH<sub>3</sub>), 4.45 (d, 1H, J=9 Hz, H-1), 4.79, 4.98 (2d, 2H, PhCH<sub>2</sub>), 5.56 (s, 1H, PhCH), 7.23-7.44 (m, 10H, aromatic protons).

Anal. Calcd for C<sub>22</sub>H<sub>26</sub>O<sub>5</sub>S : C, 65.6; H, 6.5. Found: C, 65.8; H, 6.6.

Ethyl 2-O-Acetyl-3-O-benzyl-4,6-O-benzylidene-1-thio- $\beta$ -D-glucopyranoside (11). Compound 10 (3.0 g, 7.46 mmol) was acetylated conventionally using pyridine and acetic anhydride to give 11 (3.3 g, quantitative):  $[\alpha]_D$  -29.8° (c 1.3, CHCl<sub>3</sub>); <sup>1</sup>H NMR  $\delta$  1.30 (t, 3H, SCH<sub>2</sub>CH<sub>3</sub>), 2.74 (q, 2H, SCH<sub>2</sub>CH<sub>3</sub>), 4.45 (d, 1H, J=9.5 Hz, H-1), 4.80, 4.98 (2d, 2H, PhCH<sub>2</sub>), 5.05 (t, 1H, H-2), 5.56 (s, 1H, PhCH), 7.23-7.44 (m, 10H, aromatic protons).

Anal. Calcd for C<sub>24</sub>H<sub>28</sub>O<sub>6</sub>S : C, 64.85; H, 6.35. Found : C, 64.74; H, 6.42.

Methyl 2-O-Acetyl-3-O-benzyl-4,6-O-benzylidene-β-D-glucopyranosyl-(1- $\rightarrow$ 3)-[2,3,4-tri-O-benzyl-β-L-fucopyranosyl-(1- $\rightarrow$ 2)]-4,6-di-Obenzyl-α-D-galactopyranoside (12). To a solution of 8 (1.53 g, 1.93 mmol) and 11 (1.28 g, 2.88 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (30 mL) was added MS-4Å (5 g) and the reaction was allowed to proceed in presence of NIS (977 mg, 4.34 mmol) and TfOH (33.3 µL, 0.43 mmol) as described for the preparation of 4. Column chromatography of the crude syrupy product using 10:1 toluene-Et<sub>2</sub>O then gave pure 12 (1.56 g, 69%): [α]<sub>D</sub> + 2.2° (c 1.1, CHCl<sub>3</sub>); <sup>1</sup>H NMR δ 1.22 (d, 3H, J=6.1 Hz, CCH<sub>3</sub>), 1.94 (s, 3H, COCH<sub>3</sub>), 3.37 (s, 3H, OCH<sub>3</sub>), 4.59 (d, 1H, J=9.0 Hz, H-1"), 4.77 (d, 1H, J=7.6 Hz, H-1'), 4.93 (d, 1H, J=2.4 Hz, H-1), 5.05 (t, 1H, J=8.7, H-2"), 5.53 (s, 1H, PhCH), 7.21-7.52 (m, 35H, aromatic protons).

Anal. Calcd for C<sub>70</sub>H<sub>76</sub>O<sub>16</sub>: C, 71.67; H, 6.48. Found : C, 71.48; H, 6.65.

Methyl 3-O-Benzyl-4,6-O-benzylidene-β-D-glucopyranosyl-(1→3)-[2,3,4-tri-O-benzyl-β-L-fucopyranosyl-(1→2)]-4,6-di-O-benzyl-α-D-galactopyranoside (13). A solution of 12 (1.56 g, 1.33 mmol) was treated with 0.05 M MeONa in MeOH (50 mL) as described for the preparation of 6 to give a solid mass which crystallized from EtOH yielding 13 (1.39 g, 92%): mp 165-166 °C;  $[\alpha]_D$  +3.1° (c 1.2, CHCl<sub>3</sub>); <sup>1</sup>H NMR  $\delta$  1.29 (d, 3H, J=6.3 Hz, CCH<sub>3</sub>), 3.29 (s, 3H, OCH<sub>3</sub>), 4.53 (d, 1H, J=8.7 Hz, H-1"), 4.61 (d, 1H, J=8.4 Hz, H-1'), 4.84 (d, 1H, J=3.3 Hz, H-1), 5.57 (s, 1H, PhCH), 7.20-7.50 (m, 35H, aromatic protons).

Anal. Calcd for C68H74O15 :C, 72.21; H, 6.55. Found : C, 72.04; H, 6.73.

Methyl 3-O-Benzyl-4,6-O-benzylidene-B-D-mannopyranosyl- $(1\rightarrow 3)$ -[2,3,4-tri-O-benzyl- $\beta$ -L-fucopyranosyl- $(1\rightarrow 2)$ ]-4,6-di-O-benzyl- $\alpha$ -D-galactopyranoside (14). To a solution of 13 (1.39 g, 1.23 mmol) in Me<sub>2</sub>SO (15 mL) was added 1:2 Ac<sub>2</sub>O-Me<sub>2</sub>SO (30 mL) and the mixture was stirred at room temperature for 16 h. Solvents were removed yielding the 2keto compound (1.38 g) as a yellow syrup. To a solution of this product in CH<sub>2</sub>Cl<sub>2</sub>-MeOH (80 mL, 1:1; v/v) was added NaBH<sub>4</sub> (8 g) and the mixture was stirred at 5-10 °C for 5 h. The reaction mixture was concentrated in vacuo and diluted with CH<sub>2</sub>Cl<sub>2</sub> (150 mL). The organic layer was washed successively with 5% citric acid solution, aq NaHCO<sub>3</sub>, water, dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated. Column chromatography using 4:1 toluene-Et<sub>2</sub>O gave 14 (0.86 g, 62%) together with its glucoisomer (20%): Compound 14 has  $[\alpha]_D$ +0.9° (c 1.02, CHCl<sub>3</sub>); <sup>1</sup>H NMR  $\delta$  1.26 (d, 3H, J=6.6 Hz, CCH<sub>3</sub>), 3.25 (s, 3H, OCH<sub>3</sub>), 4.65 (d, 1H, J=9.6 Hz, H-1'), 4.83 (d, 1H, J=2.7 Hz, H-1), 5.30 (bs, 1H, H-1"), 5.59 (s, 1H, PhCH), 7.19-7.50 (m, 35H, aromatic protons).

Anal. Calcd for C68H74O15: C, 72.21; H, 6.55. Found : C, 72.08; H, 6.78.

Methyl 3,4,6-Tri-O-acetyl-2-deoxy-2-phthalimido-β-D-glucopyranosyl-(1 $\rightarrow$ 2)-3-O-benzyl-4,6-O-benzylidene-β-D-mannopyranosyl -(1 $\rightarrow$ 3)-[2,3,4-tri-O-benzyl-β-L-fucopyranosyl-(1 $\rightarrow$ 2)]-4,6-di-O-benzyl-αgalactopyranoside (16). To a solution of 14 (0.86 g, 0.76 mmol) and ethyl 2,3,4-tri-O-acetyl-2-deoxy-2-phthalimido-1-thio-β-D-glucopyranoside 15 (0.5 g, 1.14 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (30 mL) was added MS-4Å (5 g) and the reaction was allowed to proceed in presence of NIS (385.2 g, 1.71 mmol) and TfOH (15.1 μL, 0.17 mmol) as described for the preparation of 4. Column chromatography using 5:1 toluene-Et<sub>2</sub>O gave 16 (0.75 g, 64%):  $[\alpha]_D - 9^\circ$  (c 0.8, CHCl<sub>3</sub>); <sup>1</sup>H NMR δ 1.13 (d, 3H, J=6.2 Hz, CCH<sub>3</sub>), 1.89, 1.94 and 1.97 (3s, 9H, 3 COCH<sub>3</sub>), 3.21 (s, 3H, OCH<sub>3</sub>), 4.62 (bs, 1H, H-1"), 4.67 (d, 1H, J=9.3 Hz, H-1'), 4.78 (d, 1H, J=3.6 Hz, H-1), 5.27 (d, 1H, J=12.3 Hz, H-1"), 5.42 (s, 1H, PhCH), 7.12-7.87 (m, 39H, aromatic protons); <sup>13</sup>C NMR δ 17.40 (CCH<sub>3</sub>), 21.14, 21.16, 21.22 (3COCH<sub>3</sub>), 55.56 (OCH<sub>3</sub>), 68.44, 68.97, 69.32 (3 C-6), 70.95, 71.12, 71.24, 72.76, 73.25, 73.67, 74.10, 74.63, 74.92, 75.19, 75.86, 77.84, 78.03, 78.16, 78.68, 79.49, 82.96, 97.38 (C-1), 101.07 (C-1"), 102.09 (C-1'), 103.45 (C-1""), 103.57 (PhCH), 123.78-140.46 (aromatic carbons), 168.27, 169.91, 170.51, 171.27, 171.32 (5 carbonyl carbons).

Anal. Calcd for C<sub>88</sub>H<sub>93</sub>O<sub>24</sub>N: C, 68.26; H, 6.01. Found: C, 68.09; H, 6.24.

Methyl 2-Acetamido-2-deoxy- $\beta$ -D-glucopyranosyl- $(1 \rightarrow 2)$ - $\beta$ -Dmannopyranosyl- $(1\rightarrow 3)$ - $[\beta$ -L-fucopyranosyl- $(1\rightarrow 2)$ ]- $\alpha$ -D-galactopyranoside (17). To a solution of 16 (700 mg, 0.45 mmol) in aqueous 95% ethanol (25 mL) was added hydrazine hydrate (2 mL). The mixture was heated at 85 °C for 2 h and then concentrated. The residue was treated with pyridine (10 mL) and  $Ac_2O$  (7 mL) at room tempture for 3 h and concentrated. The residue was purified by column chromatography using 5:1 toluene-Et<sub>2</sub>O. The product was dissolved in AcOH (5 mL) and stirred under H<sub>2</sub> for 2 days in the presence of 10% Pd-C (500 mg). The reaction mixture was then filtered through a Celite bed, concentrated, dissolved in water (2 mL), passed through a 0.45  $\mu$ m Millipore membrane and dried to afford 17 ( 211 mg, 67%):  $[\alpha]_D + 26.9^\circ$  (c 0.9, H<sub>2</sub>O); <sup>1</sup>H NMR (D<sub>2</sub>O) δ 1.11 (d, J=6.3 Hz, 3H, CCH<sub>3</sub>), 1.91 (s, 3H, NCOCH<sub>3</sub>), 3.27 (s, 3H,  $OCH_3$ ), 4.16 (bs, 1H, H-1''), 4.30 (d, 1H, J=7.8 Hz, H-1'), 4.70 (bs, 1H, H-1), 4.83 (d, 1H, J= 8.4 Hz, H-1");  $^{13}$ C NMR (D<sub>2</sub>O)  $\delta$  16.46 (CCH<sub>3</sub>), 23.16 (NCOCH<sub>3</sub>), 56.27 (OCH<sub>3</sub>), 61.64, 61.70, 62.13 (3 C-6), 67.74, 67.95, 69.46, 70.58, 70.95, 71.71, 72.61, 74.21, 76.32, 76.85, 76.96, 79.44, 99.92 (C-1), 101.17 (C-1"), 102.19 (C-1'), 103.25 (C-1""), 175.44 (NCOCH<sub>3</sub>).

Anal. Calcd for C<sub>27</sub>H<sub>47</sub>O<sub>20</sub>N: C, 45.96; H, 6.66. Found: C, 45.80; H, 6.89.

#### REFERENCES

- 1. F. Kauffmann, *The Bacteriology of Enterobacteriaceae*, Munksgaard, Copenhagen, p 19 (1966).
- 2. W. Stenzel, Zentralbl. Bakteriol. Parasitenkd. Infektionskr. Hyg., Abt. I: Orig., 243, 97 (1975).
- I. Ørskov, F. Ørskov, B. Jann and K. Jann, Bacteriol. Rev., 41, 667 (1977).
- 4. F. Ørskov and I. Ørskov, Methods Microbiol., 14, 43 (1984).
- S. R. Munford, Acta Pathol. Microbiol. Immunol. Scand., 99, 487 (1991).
- 6. A. W. Richter and R. Eby, Mol. Immunol., 22, 29 (1985).
- 7. T. Bhattarcharyya and S. Basu, Carbohydr. Res., 254, 221 (1994).
- 8. A. K. Misra, S. Basu and N. Roy, Synth. Commun., 26, 2857 (1996).
- 9. F. Kong, D. Lu and S. Zhou, Carbohydr. Res., 198, 141 (1990).
- 10. H. Lönn, Carbohydr. Res., 139, 105 (1985).
- (a) P. Konradsson, U. E. Udodong and B. Fraser-Reid, *Tetrahedron Lett.*, 31, 4313 (1990);
   (b) G. H. Veenemann, S. H. van Leeuwen and J. H. van Boom, *Tetrahedron Lett.*, 31, 1331 (1990).
- 12. P. A. Gent and R. Gigg, J. Chem. Soc., Perkin Trans. 1, 1446 (1974).
- 13. G. Zemplén, Ber. Dtsch Chem. Ges., 59, 1254 (1926).
- 14. J. S. Brimacombe, Methods Carbohydr. Chem., 6, 376 (1972).
- 15. T. Ogawa and H. Yamamoto, Agric. Biol. Chem., 49, 475 (1985).
- G. Magnusson, S. Ahlfors, J. Dahmén, K. Jansson, U. Nilsson, G. Noori, K. Stenvall and A. Tjornebo, J. Org. Chem., 55, 3932 (1990).
- 17. H. Qiu and T. B. Grindley, J. Carbohydr. Chem., 15, 95 (1996).
- M. L. Wolfrom and A. Thompson, Methods Carbohydr. Chem., 2, 211 (1982).
- 19. C. Auge, C. D. Warren and R. W. Jeanloz, *Carbohydr. Res.*, 82, 85 (1982).
- 20. H. Lönn, Carbohydr. Res., 296, 105 (1985).
- T. Ehara, A. Kameyama, Y. Yamada, H. Ishida, M. Kiso and A. Hasegawa, *Carbohydr. Res.*, 181, 237 (1996).